

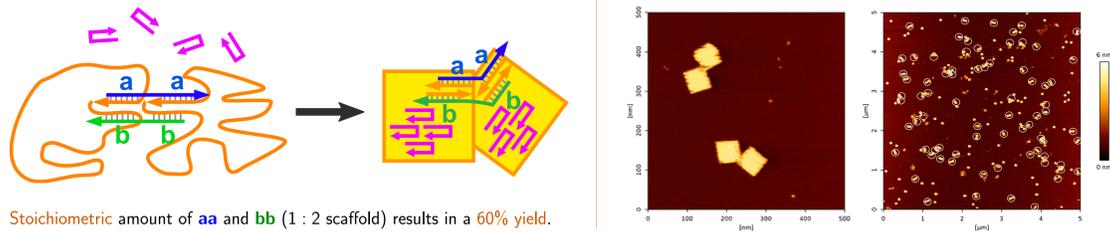
Introduction

Goal: Better understand how to build nanostructures composed of *several* DNA origamis bound together and based on *identical* scaffold strands.

Motivations:

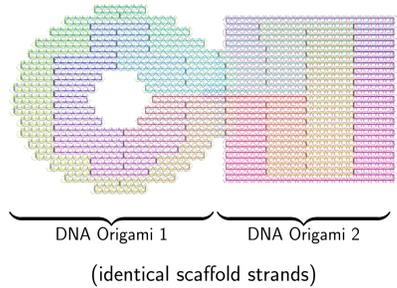
- finding a **cost-effective** method to create arbitrary objects of **larger dimension** than simple DNA origami,
- better understanding and controlling DNA origami folding pathways.

Preliminary Work: joining 2 identical origamis

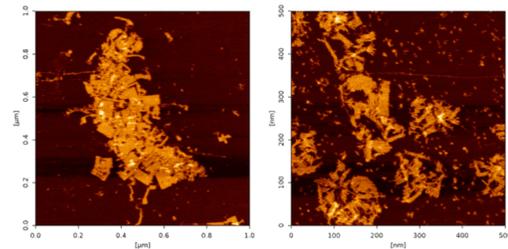


Our approach

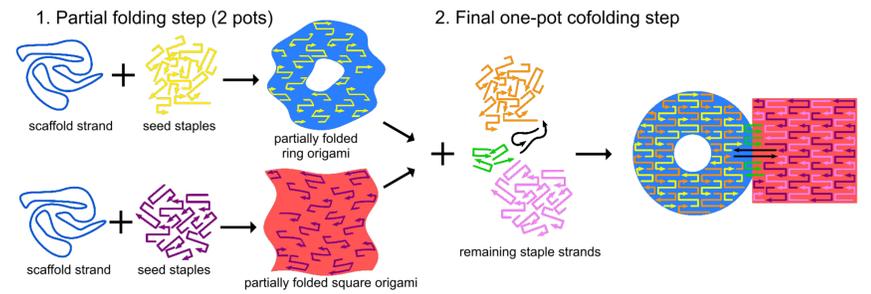
Our test design using ENSnano:



Problem: The classical **single annealing** method produces **chimeric** structures.



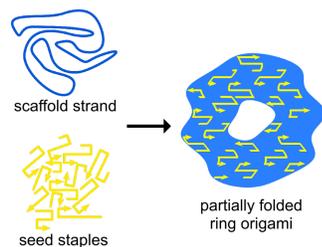
Looking for key staples using a two-stages folding method.



Key staples: looking for seed strands

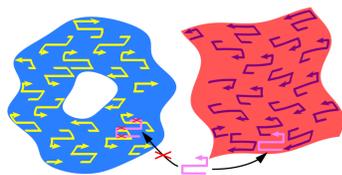
A subset of staples that:

- Pre-folds each scaffold separately (step 1),
- Is as small as possible,
- Produces the desired structure at the final annealing step (step 2).
- ⚠ Checkerboard pattern doesn't work. (see results c-d).



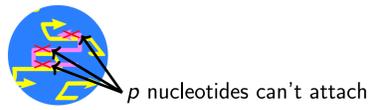
Seed strands selection method

Principle: Preventing staples from attaching to the wrong partially folded scaffold. We select **ring seed staples** so that **square staples** preferably won't attach to the partially folded **ring scaffold**.



Nucleotide counting greedy algorithm:

- **Constraint:** Uniform penalty for attaching a staple to the wrong scaffold (overlap $\geq p$ nucleotides).
- **Algorithm:** Greedily construct a small seed abiding by this constraint.



ΔG based LP algorithm:

- **Constraint:** \forall staple, $\Delta G_{\text{bad}} \geq \text{threshold}$.
- **Approximation:** ΔG is approximated as a **linear function** $\widetilde{\Delta G}$ of the binding domains:

$$\widetilde{\Delta G} \left(\begin{array}{c} \text{---} \\ \text{---} \\ \text{---} \end{array} \right) = \widetilde{\Delta G} \left(\begin{array}{c} \text{---} \\ \text{---} \end{array} \right) + \widetilde{\Delta G} \left(\begin{array}{c} \text{---} \\ \text{---} \end{array} \right) + \widetilde{\Delta G} \left(\begin{array}{c} \text{---} \\ \text{---} \end{array} \right)$$

- **Algorithm:** Solving the **integer linear program** with IBM Cplex.

Thermodynamics model

$$5' \text{---} \underset{\ast}{\text{C}} \text{---} \underset{\ast}{\text{G}} \text{---} \underset{\ast}{\text{T}} \text{---} \underset{\ast}{\text{T}} \text{---} \underset{\ast}{\text{G}} \text{---} \underset{\ast}{\text{A}} \text{---} 3'$$

$$3' \text{---} \text{G} \text{---} \text{C} \text{---} \text{A} \text{---} \text{A} \text{---} \text{C} \text{---} \text{T} \text{---} 5'$$

$\Delta G_{\text{pred}}^{\ast} = \Delta G^{\ast}(\text{CG/GC}) + \Delta G^{\ast}(\text{GT/CA}) + \Delta G^{\ast}(\text{TT/AA}) + \Delta G^{\ast}(\text{TG/AC}) + \Delta G^{\ast}(\text{GA/CT}) + \Delta G^{\ast}(\text{init.})$
(SantaLucia 1998)

Computed seed structure and experimental results

Seed algorithm	Ring seed	Square seed	2-steps co-folding AFM images	
(a) Full seed	100% (243)	100% (237)	2 × 2 μm	500 × 500 nm
(b) No seed	0% (0)	0% (0)	2 × 2 μm	1 × 1 μm
(c) Checkerboard seed (51%)	51% (124)	51% (122)	1 × 1 μm	500 × 500 nm
(d) Checkerboard seed (27%)	27% (66)	27% (63)	1 × 1 μm	500 × 500 nm
(e) 8nt-Greedy seed (48%)	46% (112)	50% (119)	2 × 2 μm	500 × 500 nm
(f) 4nt-Greedy seed (36%)	34% (84)	37% (87)	1 × 1 μm	500 × 500 nm
(g) LP 85% (48%)	47% (115)	48% (114)	500 × 500 nm	250 × 250 nm
(h) LP 70% (31%)	31% (76)	32% (75)	1 × 1 μm	500 × 500 nm

Typical folding conditions:
 Step 1: 20 nM M13mp18 scaffold strand and 100 nM staple strands in 1 × TAE buffer with 12.5 mM Mg²⁺. 95°C for 15 min, 95 → 65°C at -1°C/2 min, 65 → 45°C at -0.5°C/15 min, 45 → 25°C at -1°C/2 min.
 Step 2: Mixing tubes from step 1 and adding all staples (50 nM), 51 → 25°C at -0.5°C/min.

Conclusion

- We managed to guide multi-scaffold DNA Origami folding pathway using **properly computed small seeds**.
- Best working seed sets have **intriguing random-like structures**: *is it due to our test design?*
- Three driving forces are guiding folding pathways: **topology** (scaffold routing), **thermodynamics** (staple length and sequences), and **geometry** (the intermediate shapes of the partial assembly as it grows).
- Further analysis of obtained seed structures should help us design new experiments to better understand the relative roles of these driving forces.